

# DNA methylation, a key concept in epigenetics

**Yuanlin Wu**

Meixi Lake Middle School attached to Hunan Normal University

3394761283@qq.com

**Abstract.** DNA methylation, histone modifications, and CpG islands intricately regulate gene expression. Understanding these epigenetic mechanisms provides insights into cellular identity and how their dysregulation contributes to cancer development. Unraveling these complexities opens avenues for therapeutic interventions aimed at restoring normal epigenetic patterns in cancer cells. In light of the potential for epigenetic therapies to restore normal gene activity and enhance treatment outcomes, this work intended to present an overview of the state of knowledge about DNA methylation and its implications for cancer therapy.

**Keywords:** DNA methylation, epigenetics, DNMTs, CpG islands.

## 1. Introduction

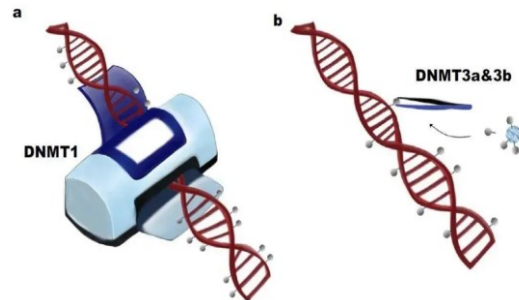
Changes in genes and epigenetics are crucial for the onset and progression of cancer. Numerous studies have been conducted on the genetic alterations that contribute to cancer, including copy number variations, deletions, insertions, and recombination events in addition to DNA missense mutations [1]. It is now widely accepted that epigenetic disruptions cause carcinogenic characteristics to accumulate as well [2], such as changes in noncoding RNAs, nucleosome location, histone alterations, and DNA methylation.

To be more specific, DNA methylation is a kind of epigenetic mechanism that includes the addition of a methyl group(CH<sub>3</sub>) into the DNA molecule. It can inhibit the expression of genes by impeding the binding of transcription factors with DNA molecules.

As an example of how DNA methylation affects the initiation or development of cancers, DNA methylation can be involved in the promoter region in the DNA sequence. While gene body DNA methylation is frequently favorably correlated with gene expression, promoter DNA methylation may be adversely correlated with the state of gene activation [3]. If the tumor-suppressor gene is methylated, it is more likely to be suppressed, thus increasing the probability of cancer. Both mechanisms contribute to gene regulation, and the whole process is highly regulated. Different mechanisms, such as somatic mutations, insertions/deletions, and genetic and epigenetic changes of DNA methylation, can independently mute or activate one of or both gene alleles, then coordinating an impact on tumor suppressor genes and oncogenes throughout the human genome [4]. What's more, it's interesting to note that somatic mutations in epigenetic modifier genes have been found in large-scale cancer genome DNA sequencing analyses. This suggests that both of genetic and epigenetic systems or mechanisms also have the ability to reinforce one another in a mutually separate way.

## 2. DNMTs - The key player of DNA methylation

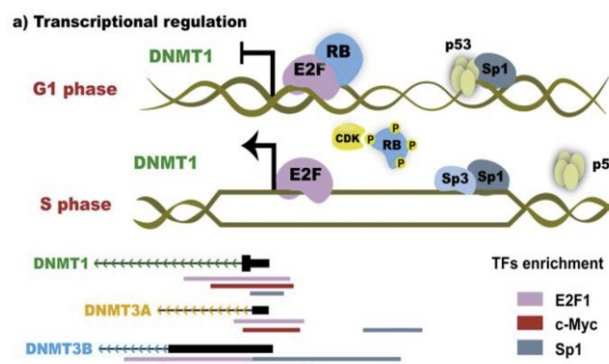
DNA methylation is the chemical alteration of S-adenosine methionine (SAM) as a CH<sub>3</sub> donor under the catalysis of DNA methyltransferases (DNMTs). It is the process of getting a methyl (CH<sub>3</sub>) group via covalent binding. It involves the addition of methyl groups to cytosine within CpG dinucleotides and is orchestrated by DNMTs, which means the DNMTs play important roles in the process of DNA methylation.



**Figure 1.** The roles of DNA methyltransferases in biochemistry.

(a) According to its definition, DNMT1 is a maintenance DNA methyltransferase. DNMT1 copies the current methylation pattern, favoring hemi-methylated DNA; (b) De novo DNMTs, such as DNMT3a and DNMT3b, transfer methyl groups donated by S-adenyl methionine (SAM) to unmethylated DNA to create new methylation patterns [33].

The primary players in this process are DNMT1, DNMT3A, and DNMT3B. DNMT1 ensures the maintenance of methylation patterns during cell division, while DNMT3A and DNMT3B contribute to de novo methylation, particularly in embryonic development.<sup>1</sup> The development of an embryo is very similar to the developing stage of cancer. Genes critical for cell cycle and cell cycle progression can be interfered with by DNMT to encourage replication. In the process of DNMT participating in DNA methylation, cytosine (C) is the most common base of methylation, and C site methylation mainly occurs on the CpG sequence, which means CpG island is also an essential point in methylation.



**Figure 2.** Transcriptional regulation and post-transcriptional modifications of DNMT1. (Adapted from Loeza-Loeaza and Beltran)

When DNMTs move from phase G1 to phase S, their expression rises. Phase G1 sees pRB dephosphorylated and bound to E2F as a result of p53's interaction with Sp1. As p53 levels fall and CDK expression rises during replication, pRB that has been phosphorylated separates from E2F and p53 separates from Sp1, which makes it easier for DNMT1 transcription to be activated. In various cell lines examined for the ENCODE project, Sp1, E2F, and c-Myc transcription factors (TFs) are enriched in the promoters of DNMT1, DNMT3A, and DNMT3B.

There are a lot of regulators that regulate the DNMTs, for example, microRNAs (miRNAs). MiRNAs are a class of small RNA molecules that play a role in regulating gene expression in cancer by targeting

specific mRNA molecules for degradation or inhibiting translation. In cancer, miRNAs are often dysregulated and can promote cancer development or act as tumor suppressor genes to suppress tumor growth. For instance, some important genetic expressions, such as the expression of tumor suppressor genes, may be restricted in the cancer cells due to unregulated DNMT expression, while this expression is carefully regulated in normal cells [30].

### 3. CpG Islands

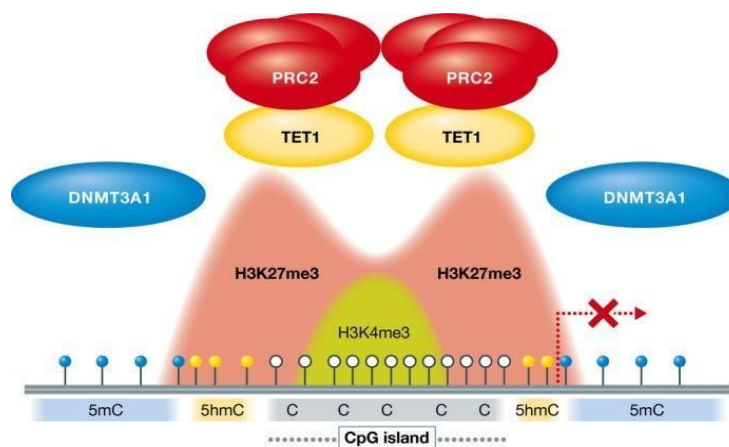
CpG islands, characterized by high CpG content, are integral to epigenetic regulation. While the majority of genomic CpGs undergo methylation, CpG islands in gene promoters typically remain unmethylated, allowing for unimpeded gene expression. So, if the CpG island is methylated, the genes that follow it will be silenced. Defined by criteria such as a sequence length of 500 base pairs and a C + G content of more than 55% [5], CpG islands serve as epigenetic signposts guiding transcriptional regulation.

The distinct methylation status of CpG islands distinguishes them as essential contributors to chromatin accessibility and gene expression control. As an illustration, CpG residues in coding areas or gene bodies in somatic cells frequently lead to active spots for mutational, as seen in the inactivation of C to T transitions in the tumor suppressor gene TP53 [5]. Unlike non-CpG island promoters, which are predominantly methylated, CpG islands play a crucial role in maintaining an open chromatin state, essential for normal cellular function. For example, promoter CpG islands are usually hypomethylated in normal cells, but hypermethylated in tumor cells [6]. This reveals the relationship between the degree of methylation in promoter CpG islands and the risk of cancers.

### 4. Histone Modifications

Histone modifications, pivotal in shaping chromatin structure, constitute another section of epigenetic regulation. These modifications, involving the addition or removal of functional groups to histone proteins, significantly impact gene expression. The intricate interplay between DNA methylation and some specific histone modifications adds complexity to the epigenetic landscape.

Promoter DNA methylation is often related with histone H3 lysine 27 trimethylation (H3K27me3), which is associated with DNMT3A1, contributing to gene silencing. (figure 1.) [16]. Conversely, gene body DNA methylation correlates with histone H3 lysine 36 trimethylation (H3K36me3), facilitating gene expression [1]. So, for instance, in the case of cancer development, if the histone H3K27me3 is methylated, the promoter is also methylated to cause the subsequent suppressor or oncogene to be silenced. DNMT3B's association with H3K36me3 emphasizes the relationship between DNA methylation and histone modifications, revealing the regulatory mechanisms that can be also applied to cancer studies.



**Figure 3.** Polycomb targeting at bivalent hypomethylated CpG islands (CGI) is influenced by DNA methylation mediated by the de novo methyltransferase DNMT3A1 [16].

Unmethylated transcriptionally inactive CGIs have been connected to methylated PRC2 targeting. Since H3K27me3 and DNA methylation can demonstrate mutual antagonism inside CGIs of mammalian genomes, CpG methylation inhibits the H3K27me3 histone mark that is catalyzed by PRC2 [29]. (PRC2, one of polycomb group proteins, which has histone methyltransferase activity, PRC2 inhibits gene expression through its histone H3K27 methyltransferase activity; TET1 is an enzyme that plays a positive role in DNA demethylation)

## **5. Dysregulation of DNA methylation**

As mentioned above, there are different types of DNA methylation, and DNA methylation can occur in different sections of DNA, so in this section we will discuss two main categories of DNA methylation, hypermethylation and hypomethylation.

### *5.1. Hypermethylation*

Hypermethylation is generally defined as an abnormal increase in the degree of methylation on DNA beyond normal levels. This state can lead to the silencing or inactivation of specific genes. In the case of cancer, we will pay particular attention to the effects of hypermethylation on tumor suppressor genes and oncogenes.

Together with genetic alterations like somatic mutations and deletions, hypermethylation in the promoter can also lead to gene silence, which is a secondary mechanism for the inactivation of the tumor suppressor genes (TSGs) [1].

On CpG island, the hypermethylation can also happen in some particular cancer species. For example, specific DNA hypermethylation of a subset, a certain small area, of the gene areas, most remarkably the concordant promoter DNA, was observed in a subset of colorectal tumors. The term CpG island methylator phenotype (CIMP) refers to this condition [1,7].

Noncoding RNA (ncRNA) is a class of RNA molecules that don't code for any proteins in cells. They perform as a role that hold the post of various significant functions in the cells, including regulation of gene expression and helping to the maintenance of chromatin structure [8]. The hypermethylation can also happen to ncRNA. For example, DNA hypermethylation of miR-127, leads to downregulation of its expression and succeeding up-regulation of the BCL6 proto-oncogene, which promotes tumorigenesis [9,10].

### *5.2. Hypomethylation*

In contrast to hypermethylation, hypomethylation is a state in which the level of methylation on DNA is lower than normal.

The majority of hypomethylation in DNA takes place in intragenic and intergenic areas. Repetitive and transposable elements, such as long interspersed nucleotide elements (LINEs) and short interspersed nucleotide elements (SINEs), are plentiful in these genomic regions [1,11]. LINE-1, can, for example, have a significant impact on gene expressions. The hypomethylation of this gene is known to be a predictor of tumor development and prognosis in a number of cancer types, including ovarian, bladder, renal, prostate, and melanoma [1].

**Table 1.** This table suggested the significance of DNA methylation in various cancers and the abundance of the dysregulation discovered across different cancers, highlighting the potential of targeting DNA methylation as a potential therapeutic strategy.

cancer type	hyper(number)	gene	hypo(number)	gene	gene(not sure)	doi
GBM(glioblastoma muliforme)					<i>IDH1</i> <i>IDH2</i> <i>O6-MGMT</i>	(Della Monica and Cuomo) <sup>17</sup> .
	1214		4353			(Ji and Zhao) <sup>18</sup> .
LGG(low-grade glioma)		<i>ARL9</i>				(Tan and Zhang) <sup>19</sup> .
					<i>HTATIP2</i>	(Dong and Deng) <sup>20</sup> .
TCGA high-grade serous ovarian tumor	168				<i>BRCA1</i>	(The Cancer Genome Atlas Research Network) <sup>21</sup> .
SKCM(skin cutaneous melanoma)	41					(Fu and Wu) <sup>22</sup> .
		<i>CLDN11</i>				
		<i>RASSF6</i>				
		<i>RASSF10</i>				
		<i>GPX3</i>				
		<i>MMP-9</i>				
		<i>SYNPO2</i>				
		<i>LINE-1</i>				
LIHC(liver hepatocellular carcinoma)	11		14			(Fan and Tu) <sup>23</sup> .
				<i>ANAPC10</i>		
				<i>ANAPC4</i>		
				<i>BUB1</i>		
				<i>AURKA</i>		
				<i>CCNB1</i>		
				<i>ROBO1</i>		
		<i>AR</i>				
		<i>CCND1</i>				
		<i>BDNF</i>				
breast cancer				<i>TSTD1</i>		(Muhamad Ansar and Thi) <sup>24</sup> .
		<i>GSTM2</i>				(Ma and Li) <sup>25</sup> .
		<i>SOD3</i>				
				<i>ADAM12</i>		
				<i>LINE-1</i>		
				<i>KCNK9</i>		
				<i>YAP1</i>		
					<i>BRCA2</i>	(Szczepanek and Skorupa) <sup>26</sup> .
				<i>RAD9</i>		
				<i>I-Oct</i>		
				<i>c-Fos</i>		
				<i>H-Ras</i>		
				<i>c-Myc</i>		
		<i>ERa</i>				
		<i>HOXA5</i>				
		<i>TMS1</i>				
UCS(uterine carcinosarcoma)	2191		365			(Li and Xing) <sup>27</sup> .
		<i>NEFM</i>				
		<i>CLEC14A</i>				
		<i>EMILIN1</i>				
					<i>MGMT</i>	(Mateusz Bujko and Kowalcwska) <sup>28</sup> .

## 6. Cancer therapy and future directions

As mentioned in the earlier section, DNMTs play a crucial role in methylation, so many studies have been conducted on DNA methyltransferase inhibitors (DNMTi).

At low dosages, DNMTis can activate genes that have been silenced, and at excessive levels, it can be harmful, or say cause cytotoxicity.<sup>12</sup> In a recent study, DNMTis can inhibit the formation of tumor endothelial cells and tumor angiogenesis,<sup>14</sup> which is a pretty interesting mechanism of the DNMTi because DNMTi can apply to any genetic modification. The huge potential of DNMTis could bring researchers some novel therapies for cancers.

DNA methyltransferase inhibitor zebularine and decitabine can significantly reduce tumor size in established tumor models by inducing re-expression and total demethylation of the p16 gene, as well as by cell cycle arrest and inducing apoptosis.<sup>13</sup> A second-generation prodrug, like guadecitabine, affects gene expression by inhibiting DNA methylation.<sup>13</sup> Guadecitabine caused dose-dependent demethylation and increase in p16 expression at both mRNA and protein level.<sup>13, 32</sup>

On the other hand, it is also explored the possibility of combination therapy with immune checkpoint inhibitors. For example, in breast cancer, the potential of DNMTi combined with PD-L1 in the treatment of breast cancer has shown significant therapeutic effect: guadecitabine with anti-PD-1/L1 immunotherapy.<sup>15</sup> By activating the anti-tumor immune response, DNMTi therapy increases MHC-I expression, attracts cytotoxic CD8<sup>+</sup>T cells, and activates the NF $\kappa$ B signaling pathway, thereby enhancing the response against PD-L1 therapy.<sup>15</sup>

## 7. Conclusion

With more and more studies looking into DNA methylation, a large amount of evidence has presented another mechanism for DNA methylation regulation. It was suggested that DNA methylation mechanisms may be regulated by microRNAs (miRNAs), and that the regulation of these genes is critical for cancer occurrence and tumor progression.<sup>30</sup> Under this circumstance, using DNMT inhibitors such as 5-aza-dC and 5-azaC could repair the expression of epi-miRNAs, a subset of miRNAs that target genes involved in epigenetic regulation.<sup>30</sup> Researchers believe targeting these EPI-miRNAs could be promising for cancer treatment.

This piece of work provides an introduction to the importance of understanding DNA methylation and its regulations, with an interest in the disruption of DNA stability in cancer. In the future researches in this field, it could be important to explain the specific mechanisms by which DNMTis interact with immune checkpoint inhibitors to enhance anti-tumor immunity. Additionally, exploring the role of other epigenetic regulators, such as histone modifiers, in combination therapies with DNMTis could provide a more comprehensive understanding of epigenetic-based cancer treatments. Moreover, investigating the potential of miRNA-based therapies, either alone or in combination with DNMTis, may offer novel approaches for cancer treatment by targeting specific epigenetic pathways.

## References

- [1] Jeltsch, A. and Jurkowska, R.Z. (2022). *DNA Methyltransferases - Role and Function*. second edition ed. Springer Nature, pp.317–348.
- [2] Hanahan, D. and Weinberg, Robert A. (2011). Hallmarks of cancer: the next Generation. *Cell*, [online] 144(5), pp.646–674. doi:<https://doi.org/10.1016/j.cell.2011.02.013>.
- [3] Liang, G. and Weisenberger, D.J. (2017). DNA methylation aberrancies as a guide for surveillance and treatment of human cancers. *Epigenetics*, 12(6), pp.416–432. doi:<https://doi.org/10.1080/15592294.2017.1311434>.
- [4] Jones, P.A. and Laird, P.W. (1999). Cancer-epigenetics comes of age. *Nature Genetics*, [online] 21(2), pp.163–167. doi:<https://doi.org/10.1038/5947>.
- [5] Jones, P.A. and Baylin, S.B. (2002). The fundamental role of epigenetic events in cancer. *Nature Reviews Genetics*, 3(6), pp.415–428. doi:<https://doi.org/10.1038/nrg816>.
- [6] Skvortsova, K., Masle-Farquhar, E. and Luu, P.-L. (2019). DNA Hypermethylation Encroachment at CpG Island Borders in Cancer Is Predisposed by H3K4 Monomethylation Patterns. *Cancer Cells*, 35(2), pp.297-314.e8. doi:<https://doi.org/10.1016/j.ccell.2019.01.004>.
- [7] Weisenberger, Daniel J., and Kimberly D. Siegmund. “CpG Island Methylator Phenotype Underlies Sporadic Microsatellite Instability and Is Tightly Associated with BRAF Mutation in Colorectal Cancer.” *Nature Genetics*, vol. 38, no. 7, 1 July 2006, pp. 787–793, [pubmed.ncbi.nlm.nih.gov/16804544/](https://pubmed.ncbi.nlm.nih.gov/16804544/), <https://doi.org/10.1038/ng1834>.
- [8] Matzke, Marjori A., and Rebecca A. Mosher. “RNA-Directed DNA Methylation: An Epigenetic Pathway of Increasing Complexity.” *Nature Reviews. Genetics*, vol. 15, no. 6, 1 June 2014, pp. 394–408, [www.ncbi.nlm.nih.gov/pubmed/24805120](https://www.ncbi.nlm.nih.gov/pubmed/24805120), <https://doi.org/10.1038/nrg3683>.
- [9] Ehrlich, Melanie. “DNA Hypomethylation in Cancer Cells.” *Epigenomics*, vol. 1, no. 2, Dec. 2009, pp. 239–259, [www.ncbi.nlm.nih.gov/pmc/articles/PMC2873040/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2873040/), <https://doi.org/10.2217/epi.09.33>.
- [10] Kulis, Marta, et al. “Epigenomic Analysis Detects Widespread Gene-Body DNA Hypomethylation in Chronic Lymphocytic Leukemia.” *Nature Genetics*, vol. 44, no. 11, 1 Nov.

- 2012, pp. 1236–1242, [www.nature.com/articles/ng.2443](http://www.nature.com/articles/ng.2443), <https://doi.org/10.1038/ng.2443>. Accessed 4 Jan. 2021.
- [11] Weiner, Alan M. “SINEs and LINEs: The Art of Biting the Hand That Feeds You.” *Current Opinion in Cell Biology*, vol. 14, no. 3, June 2002, pp. 343–350, [https://doi.org/10.1016/s0955-0674\(02\)00338-1](https://doi.org/10.1016/s0955-0674(02)00338-1). Accessed 11 Dec. 2019.
- [12] Hu, Chunhong, and Xiaohan Liu. “DNA Methyltransferase Inhibitors Combination Therapy for the Treatment of Solid Tumor: Mechanism and Clinical Application.” *Clinical Epigenetics*, vol. 13, no. 1, 27 Aug. 2021, <https://doi.org/10.1186/s13148-021-01154-x>.
- [13] Agrawal, Khushboo, and Viswanath Das. “Nucleosidic DNA Demethylating Epigenetic Drugs – a Comprehensive Review from Discovery to Clinic.” *Pharmacology & Therapeutics*, vol. 188, 1 Aug. 2018, pp. 45–79, [www.sciencedirect.com/science/article/pii/S0163725818300317](http://www.sciencedirect.com/science/article/pii/S0163725818300317), <https://doi.org/10.1016/j.pharmthera.2018.02.006>. Accessed 23 Mar. 2020.
- [14] Zhou, Shu, and Hailong Ou. “Targeting Tumor Endothelial Cells with Methyltransferase Inhibitors: Mechanisms of Action and the Potential of Combination Therapy.” *Pharmacology & Therapeutics*, vol. 247, 1 July 2023, pp. 108434–108434, <https://doi.org/10.1016/j.pharmthera.2023.108434>. Accessed 5 Mar. 2024.
- [15] Luo, Na, and Ayaka Sugiura. “Therapeutic Potential of DNA Methyltransferase Inhibitors with Immune Checkpoint Inhibitor Therapy in Breast Cancer.” *Cell Stress*, vol. 2, no. 3, 12 Mar. 2018, pp. 69–71, <https://doi.org/10.15698/cst2018.03.129>. Accessed 6 Dec. 2019.
- [16] Meehan, Richard R, and Sari Pennings. “Shoring up DNA Methylation and H3K27me3 Domain Demarcation at Developmental Genes.” *The EMBO Journal*, vol. 36, no. 23, 22 Nov. 2017, pp. 3407–3408, <https://doi.org/10.15252/embj.201798498>. Accessed 12 Dec. 2022.
- [17] Dea Monica, Rosa, and Mariella Cuomo. “MGMT and Whole-Genome DNA Methylation Impacts on Diagnosis, Prognosis and Therapy of Glioblastoma Multiforme.” *International Journal of Molecular Sciences*, vol. 23, no. 13, 1 Jan. 2022, p. 7148, [www.mdpi.com/1422-0067/23/13/7148](http://www.mdpi.com/1422-0067/23/13/7148), <https://doi.org/10.3390/ijms23137148>. Accessed 24 Mar. 2023.
- [18] Ji, Jianghuai, and Lei Zhao. “Genome-Wide DNA Methylation Regulation Analysis of Long Non-Coding RNAs in Glioblastoma.” *International Journal of Molecular Medicine*, 15 Apr. 2020, <https://doi.org/10.3892/ijmm.2020.4579>. Accessed 8 May 2022.
- [19] Tan, Yutang, and Suojun Zhang. “Prognostic Significance of ARL9 and Its Methylation in Low-Grade Glioma.” *Genomics*, vol. 112, no. 6, 1 Nov. 2020, pp. 4808–4816, <https://doi.org/10.1016/j.ygeno.2020.08.035>. Accessed 22 Oct. 2023.
- [20] Dong, Xingyu, and Qingshan Deng. “Downregulation of HTATIP2 Expression Is Associated with Promoter Methylation and Poor Prognosis in Glioma.” *Experimental and Molecular Pathology*, vol. 98, no. 2, 1 Apr. 2015, pp. 192–199, <https://doi.org/10.1016/j.yexmp.2015.01.013>. Accessed 1 June 2020.
- [21] The Cancer Genome Atlas Research Network. “Integrated Genomic Analyses of Ovarian Carcinoma.” *Nature*, vol. 474, no. 7353, June 2011, pp. 609–615, <https://doi.org/10.1038/nature10166>.
- [22] Fu, Siqi, and Haijing Wu. “DNA Methylation/Hydroxymethylation in Melanoma.” *Oncotarget*, vol. 8, no. 44, 30 May 2017, pp. 78163–78173, <https://doi.org/10.18632/oncotarget.18293>. Accessed 5 June 2020.
- [23] Fan, Guorun, and Yaqin Tu. “DNA Methylation Biomarkers for Hepatocellular Carcinoma.” *Cancer Cell International*, vol. 18, no. 1, 17 Sept. 2018, <https://doi.org/10.1186/s12935-018-0629-5>.
- [24] Muhamad Ansar, and Le Thi. “Promoter Hypomethylation and Overexpression of TSTD1 Mediate Poor Treatment Response in Breast Cancer.” *Frontiers in Oncology*, vol. 12, 7 Nov. 2022, <https://doi.org/10.3389/fonc.2022.1004261>. Accessed 16 Mar. 2024.
- [25] Ma, Lingyuan, and Chenyu Li. “The Mechanism of DNA Methylation and MiRNA in Breast Cancer.” *International Journal of Molecular Sciences*, vol. 24, no. 11, 1 Jan. 2023, p. 9360,

- www.mdpi.com/1422-0067/24/11/9360, <https://doi.org/10.3390/ijms24119360>. Accessed 10 Aug. 2023.
- [26] Szczepanek, Joanna, and Monika Skorupa. “Harnessing Epigenetics for Breast Cancer Therapy: The Role of DNA Methylation, Histone Modifications, and MicroRNA.” *International Journal of Molecular Sciences*, vol. 24, no. 8, 13 Apr. 2023, p. 7235, pubmed.ncbi.nlm.nih.gov/37108398/, <https://doi.org/10.3390/ijms24087235>.
  - [27] Li, Jing, and Xiaoyun Xing. “Whole-Genome DNA Methylation Profiling Identifies Epigenetic Signatures of Uterine Carcinosarcoma.” *Neoplasia*, vol. 19, no. 2, Feb. 2017, pp. 100–111, <https://doi.org/10.1016/j.neo.2016.12.009>. Accessed 30 Mar. 2020.
  - [28] Mateusz Bujko, and Magdalena Kowalewska. “The Promoter Methylation and Expression of the O6-Methylguanine-DNA Methyltransferase Gene in Uterine Sarcoma and Carcinosarcoma.” *Oncology Letters*, vol. 4, no. 3, 22 June 2012, pp. 551–555, <https://doi.org/10.3892/ol.2012.771>. Accessed 16 Mar. 2024.
  - [29] Manzo, Massimiliano, and Joël Wirz. “Isoform-Specific Localization of DNMT3A Regulates DNA Methylation Fidelity at Bivalent CpG Islands.” *The EMBO Journal*, vol. 36, no. 23, 26 Oct. 2017, pp. 3421–3434, <https://doi.org/10.15252/embj.201797038>. Accessed 20 Sept. 2023.
  - [30] Karimzadeh, Mohammad Reza, and Peyman Pourdavoud. “Regulation of DNA Methylation Machinery by Epi-MiRNAs in Human Cancer: Emerging New Targets in Cancer Therapy.” *Cancer Gene Therapy*, 10 Aug. 2020, <https://doi.org/10.1038/s41417-020-00210-7>. Accessed 3 Mar. 2021.
  - [31] Loeza-Loeza, Jaqueline, and Adriana S. Beltran. “DNMTs and Impact of CpG Content, Transcription Factors, Consensus Motifs, LncRNAs, and Histone Marks on DNA Methylation.” *Genes*, vol. 11, no. 11, 12 Nov. 2020, p. 1336, <https://doi.org/10.3390/genes11111336>. Accessed 3 Mar. 2021.
  - [32] Yoo, Christine B, and Shinwu Jeong. Delivery of 5-Aza-2'-Deoxycytidine to Cells Using Oligodeoxynucleotides. Vol. 67, no. 13, 1 July 2007, pp. 6400–6408, <https://doi.org/10.1158/0008-5472.can-07-0251>. Accessed 25 June 2023.
  - [33] Cui, Di, and Xiangru Xu. “DNA Methyltransferases, DNA Methylation, and Age-Associated Cognitive Function.” *International Journal of Molecular Sciences*, vol. 19, no. 5, 28 Apr. 2018, p. 1315, <https://doi.org/10.3390/ijms19051315>. Accessed 7 Nov. 2019.